

# Association of fat mass profile with natriuretic peptide receptor alpha in subcutaneous adipose tissue of medication-free healthy men: A cross-sectional study

**[version 1; referees: 1 approved with reservations]**

Petros C. Dinas<sup>12</sup>

Conceptualization

Data Curation

Formal Analysis

Investigation

Resources

Writing – Original Draft Preparation

URI: <https://orcid.org/0000-0001-6853-9238>

, Eleni Nintou<sup>1</sup>

Conceptualization

Writing – Original Draft Preparation

Writing – Review & Editing

, Dimitra Psychou<sup>3</sup>

Investigation

Resources

Writing – Review & Editing

, Marnie Granzotto<sup>4</sup>

Methodology

Resources

Writing – Review & Editing

, Marco Rossato<sup>4</sup>

Methodology

Resources

Writing – Review & Editing

, Roberto Vettor<sup>4</sup>

Methodology

Resources

Writing – Review & Editing  
, Athanasios Z. Jamurtas<sup>3</sup>

Conceptualization

Resources

Writing – Review & Editing  
, Yiannis Koutedakis<sup>23</sup>

Formal Analysis

Supervision

Writing – Review & Editing

URI: <https://orcid.org/0000-0002-7065-9447>  
, George S. Metsios<sup>2</sup>

Formal Analysis

Supervision

Writing – Review & Editing  
, Andreas D. Flouris<sup>a1</sup>

Conceptualization

Funding Acquisition

Supervision

Writing – Review & Editing

URI: <https://orcid.org/0000-0002-9823-3915>

[1] FAME Laboratory, Department of Exercise Science, University of Thessaly, Trikala, Greece

[2] Institute of Sport, Faculty of Education Health and Wellbeing, University of Wolverhampton, Walsall, UK

[3] School of Physical Education and Exercise Science, University of Thessaly, Trikala, Greece

[4] Department of Medicine – DIMED, Internal Medicine 3, University of Padova, Padova , Italy

Author notes:

Correspondence to: [a] [andreasflouris@gmail.com](mailto:andreasflouris@gmail.com)

No competing interests were disclosed.

**Background:** Atrial natriuretic peptide increases lipolysis in human adipocytes by binding to natriuretic peptide receptor-A (NPRA). The aim of the current study was to examine the associations of NPRA mRNA of subcutaneous adipose tissue with fat mass, fat-free mass, body mass index (BMI) and arterial blood pressure in medication-free healthy men.

**Method:** Thirty-two volunteers [age (years):  $36.06 \pm 7.36$ , BMI:  $27.60 \pm 4.63$  ( $\text{kg}/\text{m}^2$ )] underwent assessments of body height/weight, % fat mass, fat-free mass (kg), blood pressure, and a subcutaneous adipose tissue biopsy via a surgical technique.

## Abstract

**Results:** We found that NPRA mRNA was negatively associated with % fat mass ( $r = -0.40$ ,  $R^2 = 0.16$ ,  $p = 0.03$ ) and BMI ( $r = -0.45$ ,  $R^2 = 0.20$ ,  $p = 0.01$ ). Cohen's  $f^2$  effect size analyses showed a small effect size between NPRA mRNA and BMI ( $f^2 = 0.25$ ). One-way analysis of variance with Bonferroni post-hoc tests showed a tendency for mean differences of NPRA mRNA across BMI categories ( $p = 0.06$ ). This was confirmed by Cohen's  $d$  effect size analyses revealing a large effect size of NPRA mRNA between obese individuals ( $\text{BMI} \geq 30 \text{ kg}/\text{m}^2$ ) and either normal weight ( $\text{BMI} = 19\text{--}25 \text{ kg}/\text{m}^2$ ;  $d = 0.94$ ) or overweight ( $\text{BMI} = 25\text{--}30 \text{ kg}/\text{m}^2$ ;  $d = 1.12$ ) individuals.

**Conclusions:** NPRA mRNA is negatively associated with % fat mass and BMI in medication-free healthy men, suggesting a possible role of NPRA in the control of fat mass accumulation.

## Introduction

Atrial natriuretic peptide (ANP) lowers arterial pressure to maintain fluid volume homeostasis, thus protecting against renal and cardiac pathogenesis<sup>1</sup>. ANP also increases lipolysis in human adipocytes<sup>2</sup> by binding to natriuretic peptide receptor-A (NPRA)<sup>3</sup>. NPRA is less expressed in subcutaneous adipose tissue (SAT) in obese individuals and type 2 diabetes patients than in normal glucose tolerant individuals<sup>4</sup>. Also, NPRA signalling in skeletal muscle may enhance long-term insulin sensitivity<sup>5</sup>. Collectively, NPRA may potentially treat obesity-related disorders while ANP may play a role in the therapeutic mechanisms of beta-adrenoceptor antagonists in the mitigation of heart dysfunction and utilization of lipid mobilization<sup>6</sup>. However, the role of ANP in lipolysis has been primarily investigated mainly *in vitro* models<sup>7–9</sup>, in human blood cells from individuals under medication treatment<sup>10</sup>, and in animal models<sup>9</sup>. To our knowledge, no such information is currently available in relation to the role of its receptor (NPRA) on the adipocytes of healthy individuals. Therefore, the aim of the current study was to examine the associations of NPRA mRNA of SAT with fat mass, fat-free mass (FFM), body mass index (BMI) and arterial blood pressure (BP) in medication-free healthy men.

## Methods

The study was approved by the Ethics Committee of the University of Thessaly (protocol no. 698/2013). The inclusion criteria were: healthy adult men, non-smokers, no chronic disease and/or being under medication treatment. The participants were recruited by advertisements in a local newspaper in Trikala, Thessaly, Greece and the data collection started in July 2013

and ended in June 2014. Written consent was obtained from the 32 healthy men recruited [age (years):  $36.06 \pm 7.36$ , BMI:  $27.60 \pm 4.63$  ( $\text{kg}/\text{m}^2$ )].

To avoid misleading results, the participants refrained from exercise, alcohol, and passive smoking 72h prior the measurements, while they followed an overnight fast before they visit the physiology laboratory in the School of Exercise Science between 07:00–09:00 am. PCD and DP performed the following measurements: body height using a Seca 220 (Hamburg, Germany) stadiometer, body weight using a precision scale (KERN & Sohn GmbH, Version 5.3, Germany) and blood pressure (BP) using a Standard Aneroid sphygmomanometer (Medisave, UK) according to standard guidelines<sup>24</sup>. Briefly, participants were instructed to empty their bladders and seat for five minutes in a relaxed back rest position without speaking. BP readings were taken twice, each two minutes apart, while the mean of the two BP readings was considered as the final BP values. Fat mass percentage (%FM) and FFM were measured via bioelectrical impedance using a body composition monitor (Fresenius Medical Care AG & Co. KGaA D-61346 Bad Hamburg, Germany).

Following the aforementioned measurements, the participants underwent a SAT biopsy in the physiology laboratory by a trained physician, as previously described<sup>11</sup>. Briefly, the site of the incision was disinfected and a 10 ml of 2% xylocaine (no adrenaline) was injected for local anaesthesia. An incision of 2–2.5 cm was made 3–5 cm to the left of the navel. Nearly 500 mg of subcutaneous adipose tissue was captured and removed. The NPRA mRNA analysis of SAT samples is described elsewhere<sup>12</sup>. Briefly, total RNA was extracted using RNeasy Lipid Tissue mini kit (QIAGEN). First-strand cDNAs were synthesized from equal amounts of total RNA using random primers and M-MLV reverse transcriptase (Promega). Quantitative real-time polymerase chain reaction was performed using Sybr Green fluorophore. 18S rRNA gene was used as a reference for normalization.

Following previous methodology, we removed two mean values (i.e. outliers) of NPRA mRNA that were at a distance of more than two standard deviations from the mean of the distribution<sup>13, 14</sup>. Also, there were three missing values in the NPRA mRNA analysis of SAT samples due to failure to extract RNA from adipose tissue. Eventually, 27 NPRA mRNA values were included in the statistical analysis using SPSS (version 24; SPSS Inc., Chicago, IL, USA). Normal distribution was determined using Shapiro-Wilk test, whereas Pearson's correlation coefficient, linear regression, and Cohen's  $f^2$  effect size ( $R^2/1-R^2$ )<sup>15</sup> were used to detect associations between NPRA mRNA, %FM, FFM (kg), BMI, and BP. We also used one-way analysis of variance (ANOVA) with Bonferroni post-hoc tests, and Cohen's  $d$  effect size analyses to explore the mean differences of NPRA mRNA across different BMI categories [normal weight  $<25 \text{ kg}/\text{m}^2$  ( $n=9$ ); overweight  $25\text{--}30 \text{ kg}/\text{m}^2$  ( $n=9$ ); obese  $>30 \text{ kg}/\text{m}^2$  ( $n=9$ )]. The level of significance was set at  $p \leq 0.05$ .

## Results

The participants' characteristics are provided in Table 1. The NPRA mRNA was negatively correlated with %FM ( $r=-0.40$ ,  $p=0.03$ ) (Figure 1) and BMI ( $r=-0.45$ ,  $p=0.01$ ) (Figure 2). No associations were found between NPRA mRNA and FFM, systolic or diastolic BP ( $p>0.05$ ).

Linear regression analyses confirmed the associations between NPRA mRNA and %FM ( $R^2=0.16$ ,  $p=0.03$ ) as well as BMI ( $R^2=0.20$ ,  $p=0.01$ ). Cohen's  $f^2$  effect size analyses showed a small effect size between NPRA mRNA and BMI ( $f^2=0.25$ ), however, no effect size was detected between NPRA mRNA and %FM ( $f^2<0.20$ ). ANOVA demonstrated a strong tendency for mean differences in NPRA mRNA across BMI categories ( $p=0.06$ ). This was

confirmed by Cohen's *d* effect size analyses in NPRA mRNA, revealing large effect sizes between obese individuals ( $\text{BMI} \geq 30 \text{ kg/m}^2$ ) and either normal weight ( $\text{BMI} < 25 \text{ kg/m}^2$ ;  $d=0.94$ ) or overweight ( $\text{BMI}=25\text{--}30 \text{ kg/m}^2$ ;  $d=1.12$ ) individuals.

Table 1. Characteristics of the participants.

Age (years) (n=32)	36.06±7.36
BMI ( $\text{kg/m}^2$ ) (n=32)	27.60±4.62
Fat mass (%) (n=32)	28.32±8.87
Fat free mass (kg) (n=32)	52.90±5.02
Systolic blood pressure (mmHg) (n=32)	124.28±10.09
Diastolic blood pressure (mmHg) (n=32)	84.28±6.91

[i] BMI: Body mass index

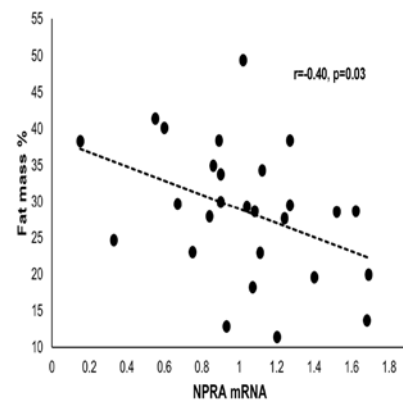


Figure 1. Association of NPRA mRNA with fat mass %.

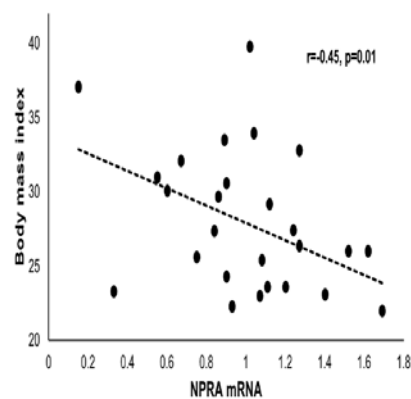


Figure 2. Association of NPRA mRNA with body mass index.

Dataset 1. Subcutaneous adipose tissue NPRA mRNA of medication-free healthy men.

BMI=Body mass index; SBP=Systolic blood pressure; DBP=Diastolic blood pressure; FFM=Fat-free mass; NPRA= Natriuretic peptide receptor-A. BMI categories= 1.  $<25 \text{ kg/m}^2$ , 2.  $25\text{--}30 \text{ kg/m}^2$ , 3.  $>30 \text{ kg/m}^2$ .

[Click here](#) to access the data.

[10.5256/f1000research.14198.d197694](https://doi.org/10.5256/f1000research.14198.d197694)

## Discussion and conclusions

We have shown that the NPRA mRNA is negatively associated with %FM and BMI in medication-free healthy men and that it is expressed less in obese compared to lean individuals. Previous evidence showed that NPRA mRNA is expressed less in normal glucose tolerant individuals than in type 2 diabetes patients<sup>4</sup>, while it is positively associated with insulin sensitivity<sup>4</sup>. Given that insulin sensitivity is negatively associated with excessive FM in humans<sup>16,17</sup> the inverse association of NPRA mRNA with %FM and BMI observed in the current study suggests a possible role of NPRA in lowering FM in humans. Indeed, it has been established that natriuretic peptides by binding to NPRA, increase cyclic guanosine monophosphate – a well-known intracellular second messenger – which phosphorylates protein kinase G leading to activation of hormone sensitive lipase<sup>18,19</sup>. This process mediates triglyceride degradation (i.e. lipolysis), which subsequently increases fatty acid availability<sup>18</sup>. Furthermore, findings in mice showed that the lack of NPRA gene may increase FM<sup>9</sup>. Also, NPRA signalling as part of ANP/NPRA axis may induce a browning of white adipocytes, indicates increased energy expenditure and thus, a potential to lessen obesity<sup>30</sup>.

The current study may be affected by methodological limitations such as the lack of insulin sensitivity measurements and *a priori* power calculation to determine the sample size. However, a post-measurements power calculation was conducted using an online software (DSS Research) to test statistical power. This revealed 89% of statistical power for the 27 available samples, based on the NPRA mRNA value ( $1.02 \pm 0.38$ ) we detected in our study and expected NPRA mRNA value ( $0.81 \pm 0.08$ ) from a previous similar study that examined NPRA in SAT in humans<sup>4</sup>. Another limitation was the lack of triglyceride, cholesterol and ANP plasma levels, to determine whether there is an association with NPRA mRNA, given the association of the aforementioned factors with lipolysis<sup>2,18</sup>. Previous evidence also suggests that, large adipocytes express higher NPRA mRNA than small adipocytes, indicates enhanced ANP-stimulated lipolysis in large adipocytes<sup>25</sup>. However, we include no analysis to determine the size of the examined adipocytes and its possible association with NPRA mRNA. Furthermore, brain natriuretic peptide (BNP) may alter expression of NPRA to release free fatty acids from adipose tissue, while obesity is inversely associated with circulating BNP, a situation known as “natriuretic handicap”<sup>26,27</sup>. Nevertheless, circulating BNP was not measured in our study to examine whether there is an association with NPRA mRNA. Also, ANP may inhibit the secretion of adipokines and cytokines from adipose tissue, thus may decrease chronic inflammation and insulin resistance<sup>28,29</sup>, however, ANP levels were not determined in our study. It would also be helpful to determine hormone sensitive lipase in the current study, given the action of ANP/BNP on lipolytic hormones<sup>2,6</sup>. Finally, the current results are limited to the Greek population and male participants, therefore, they should be treated with caution when apply to other ethnicities and females.

In conclusion, NPRA mRNA is negatively associated with %FM and BMI in medication-free healthy men, suggesting a possible role of ANP/NPRA axis in the control of FM accumulation. To date, the investigation of NPRA has mainly focused either on circulating and muscle NPRA <sup>20–22</sup> or on medication-dependent NPRA measurements <sup>4, 10</sup>. Our study indicates that NPRA may also play role in FM profile of healthy individuals, which should be further explored in a cause-and-effect research setting.

### Data availability

Dataset 1: Subcutaneous adipose tissue NPRA mRNA of medication-free healthy men 10.5256/f1000research.14198.d197694 <sup>23</sup>

BMI=Body mass index; SBP=Systolic blood pressure; DBP=Diastolic blood pressure; FFM=Fat-free mass; NPRA= Natriuretic peptide receptor-A.

BMI categories= 1. <25 kg/m<sup>2</sup>, 2. 25–30 kg/m<sup>2</sup>, 3. >30 kg/m<sup>2</sup>.

### Competing interests

No competing interests were disclosed.

### Grant information

This study was supported by funding from the European Union 7<sup>th</sup> Framework Program (FP7-PEOPLE-2012-IRSES grant no. 319010; FP7-PEOPLE-2013-IRSES grant no. 612547).

*The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

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